

EFFECTS STRESSORS ON SURVIVAL AND DAM PASSAGE OF JUVENILE CHINOOK SALMON: IMPLICATIONS FOR JUVENILE FISH PASSAGE

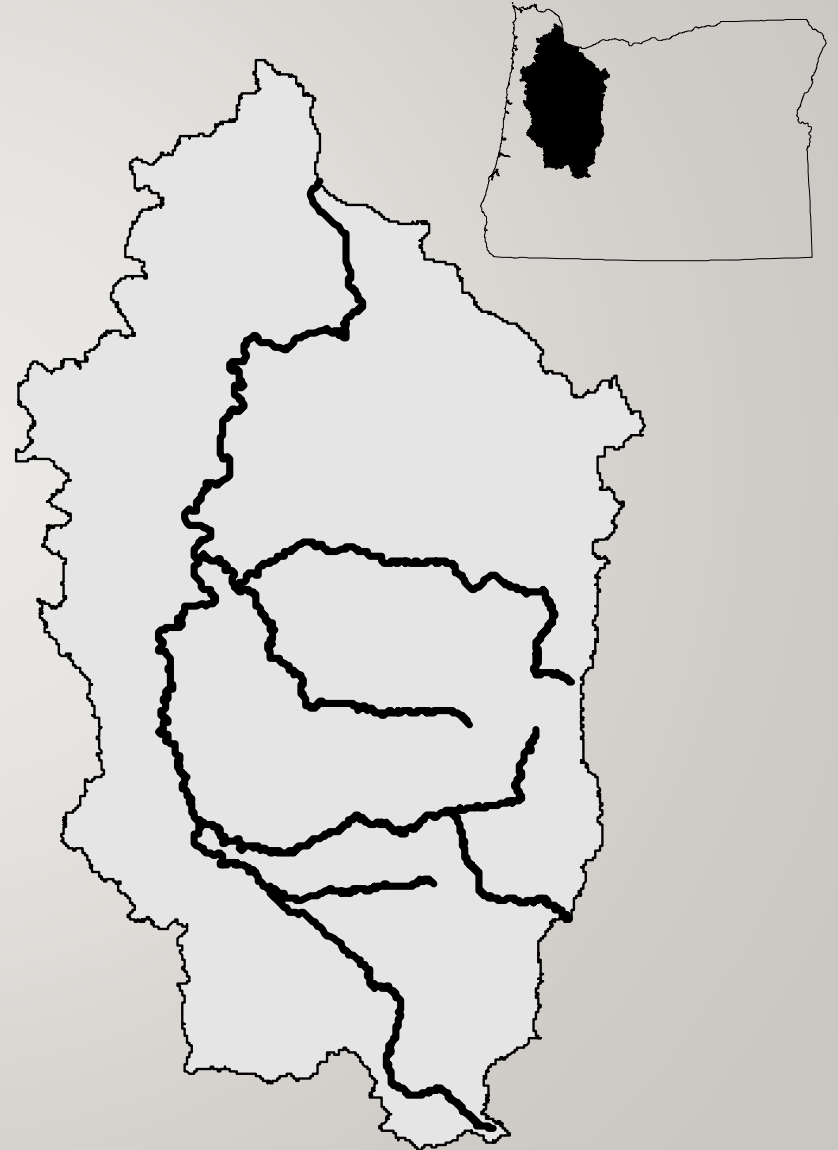
JAMES PETERSON, TRAVIS NEAL, MICHAEL KENT, JUSTIN SANDERS, AND CARL SCHRECK



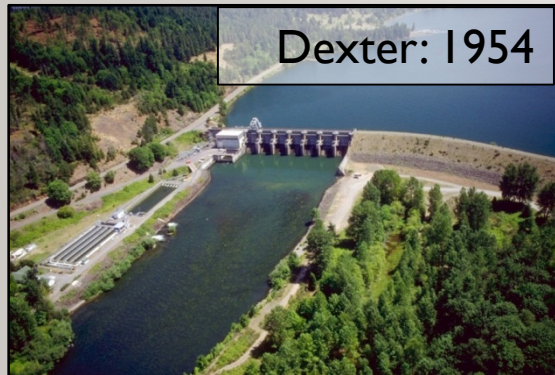
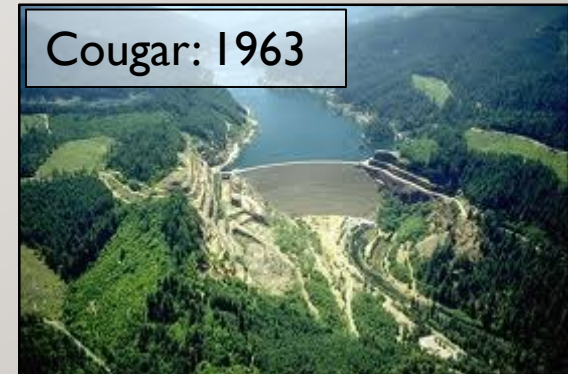
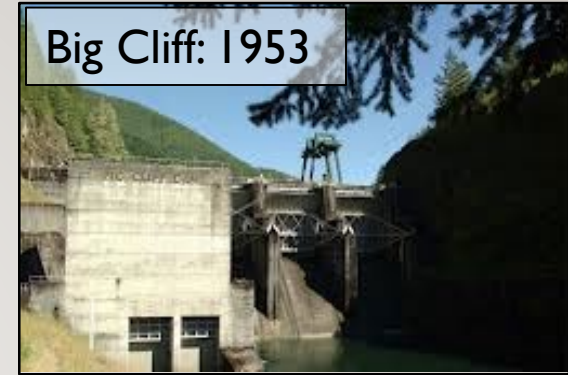
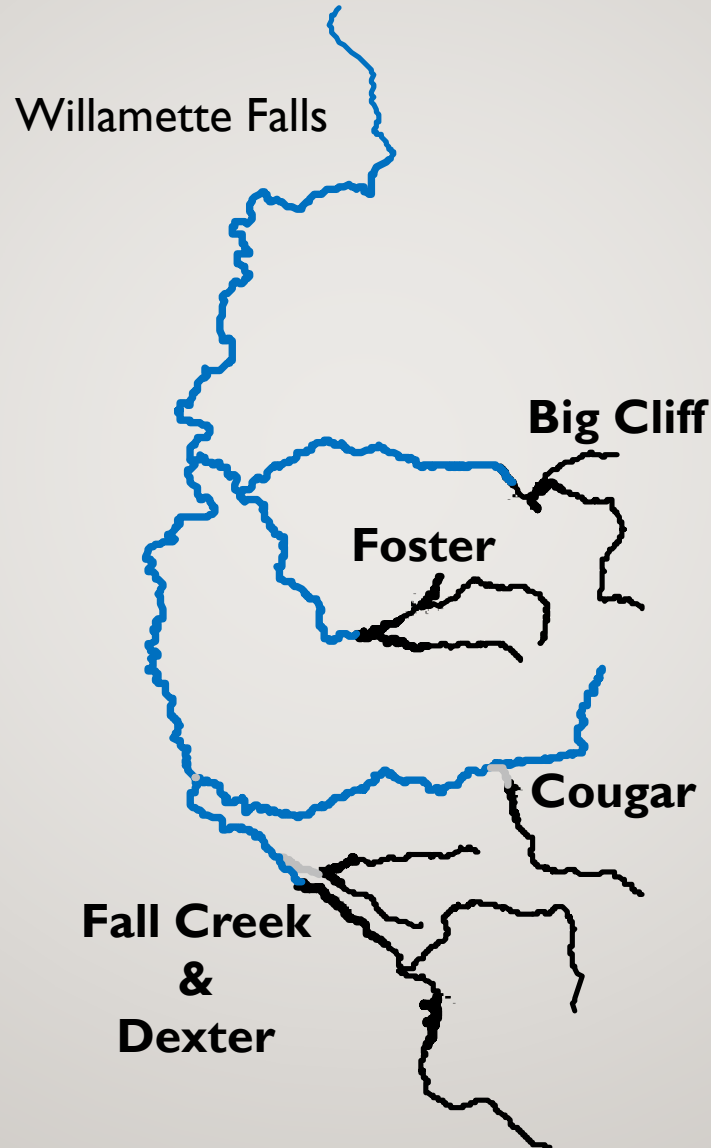
WILLAMETTE BASIN SPRING CHINOOK

Anadromous species of conservation need

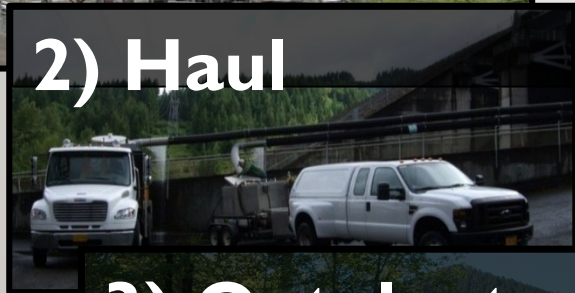
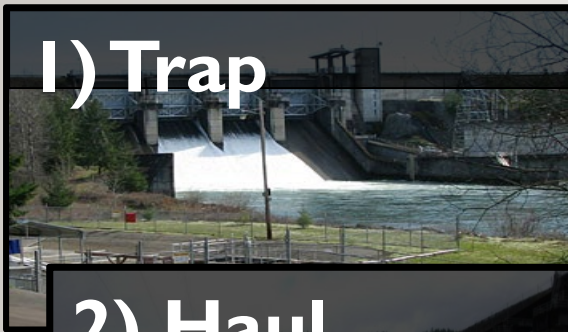
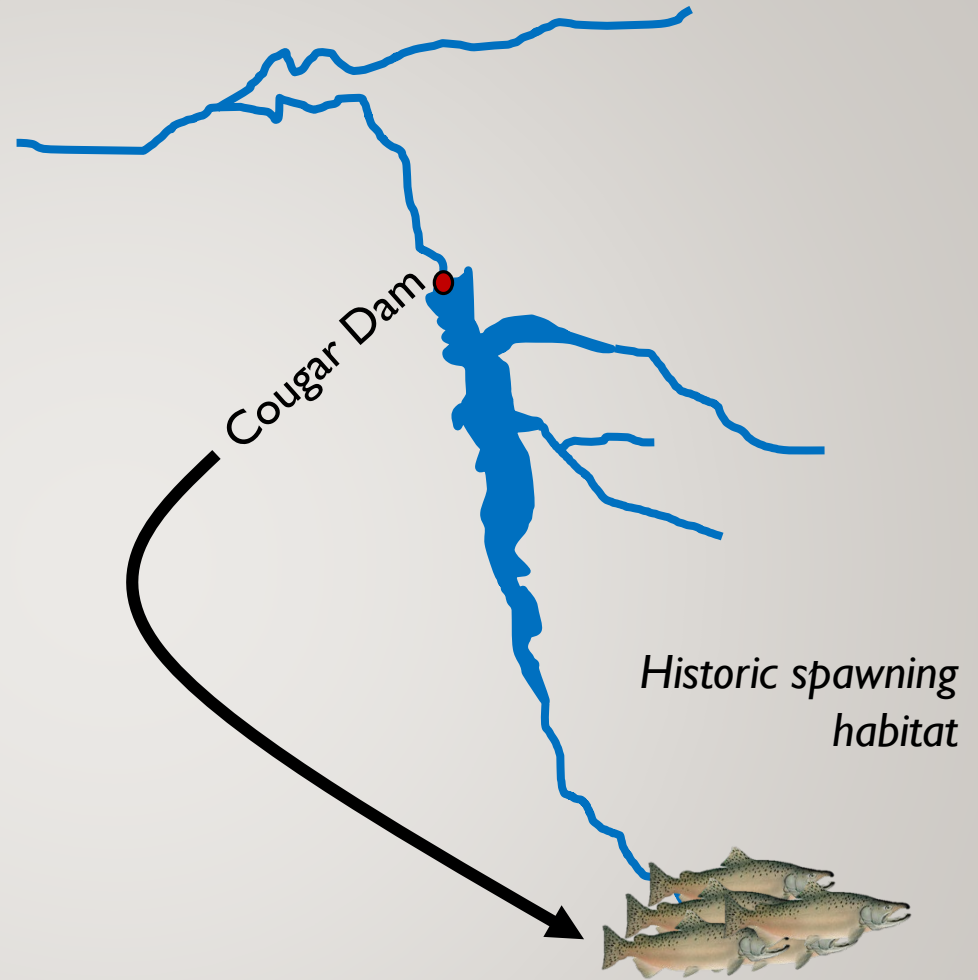
- Threatened status 1999**
- Anthropogenic modifications**



1950-60s BARRIERS TO ADULT MIGRATION



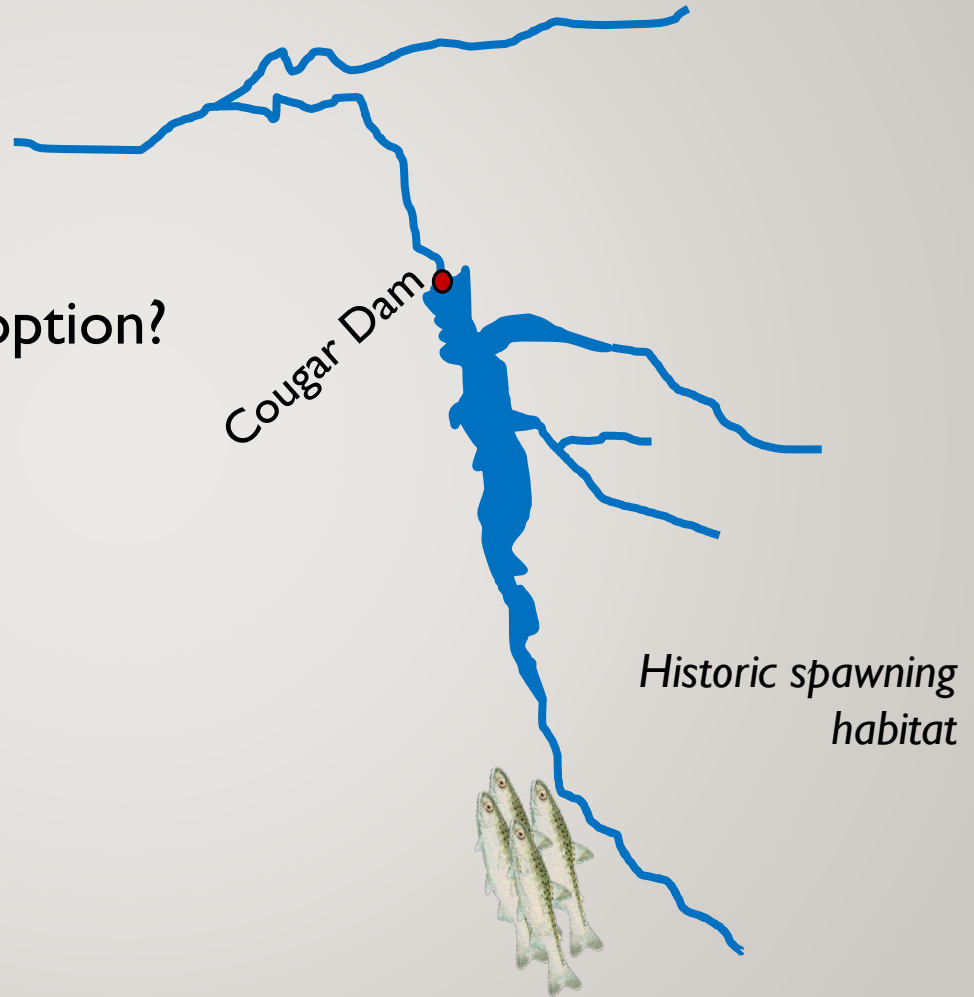
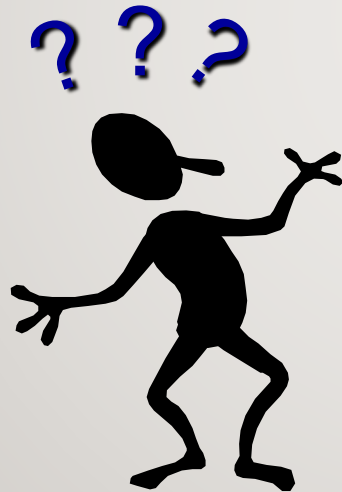
REINTRODUCTION ABOVE DAMS



NATURAL PRODUCTION!

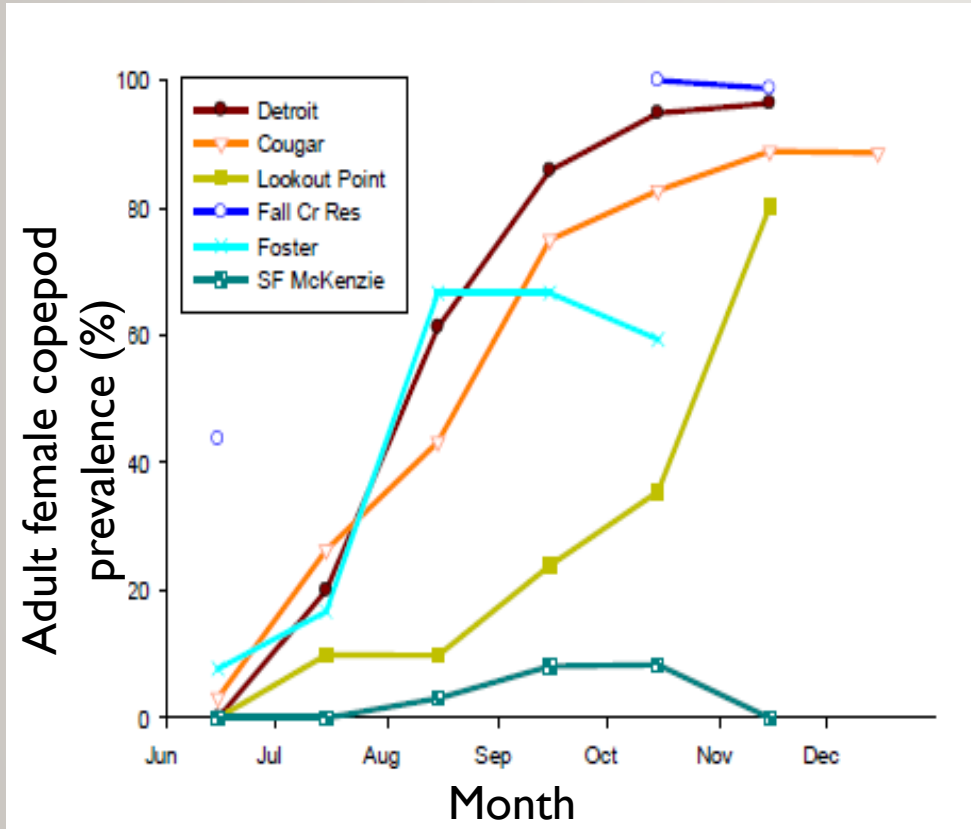
But....

What's the best passage option?



A COMPLICATION!!

Parasitic copepodids



COPEPODS AFFECT FISH PERFORMANCE

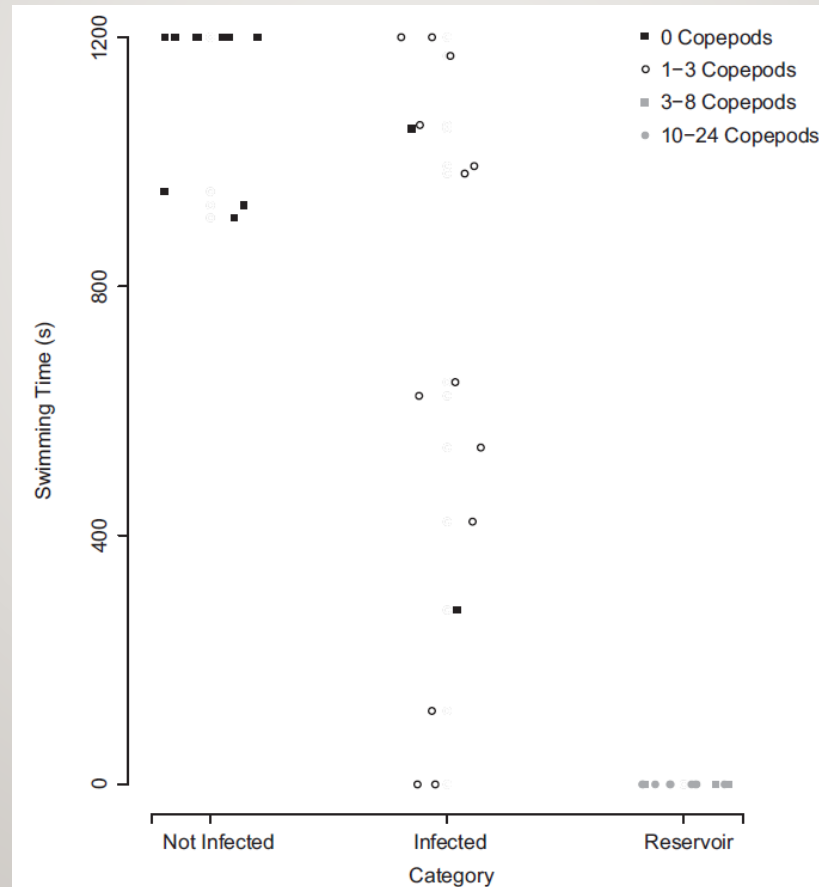
“Trap & Haul” with Copepods

- Cougar, natural infection trap & ~ 2-3 hr Haul
 - 34 dead out of 40 in 5 days
- Lab infected, netting & bucket transfer ~20 sec
 - 2 dead out of 40 in 2 days, 0 uninfected dead
- Lab infected, netting & bucket transfer ~20 sec
 - 2 dead out of 63 in 2 days, 0 uninfected dead
- Lab infected, netting & IP injection ~ 1 min
 - 30 dead out of 30 in 10 days, 0 uninfected dead

COPEPODS AFFECT FISH PERFORMANCE

Post release performance with Copepods

- Even lightly infected fish have greatly reduced swimming capability



OBJECTIVES

Ultimate goal: *safe and effective downstream juvenile Chinook passage (salmon recovery)*

Integrated approach

Laboratory studies

Infection

Fish performance

Stress

Field research

Spatial temporal distribution

Seasonality

Dynamics

Modeling

Decision Analysis



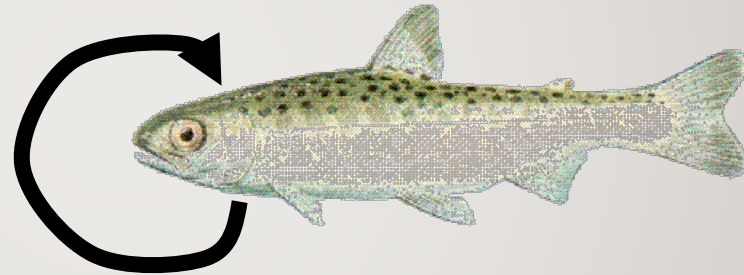
BUT FIRST....

Definitions:

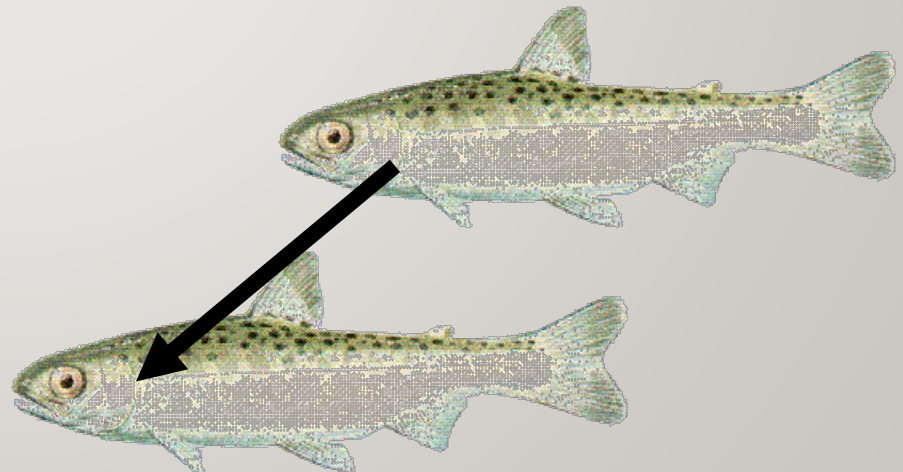
Infection rate (prevalence) = proportion of infected fish

Infection intensity = number copepods per infected fish

Autoinfection



Cross infection



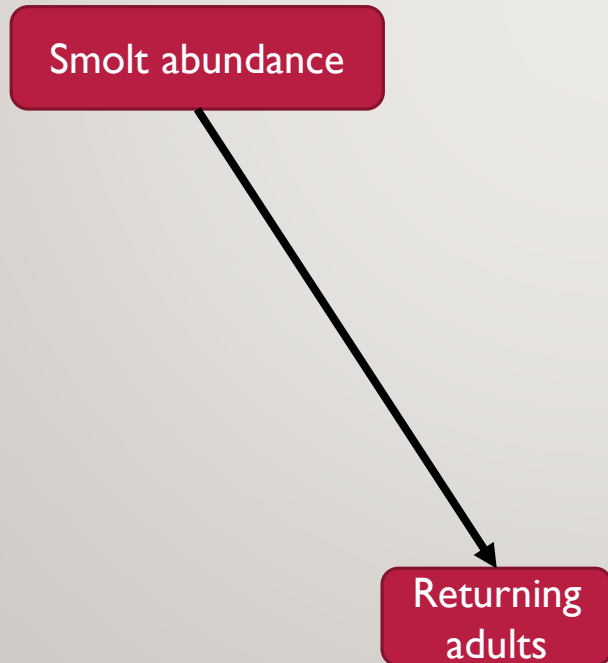
INTEGRATED APPROACH?

Our fundamental objective

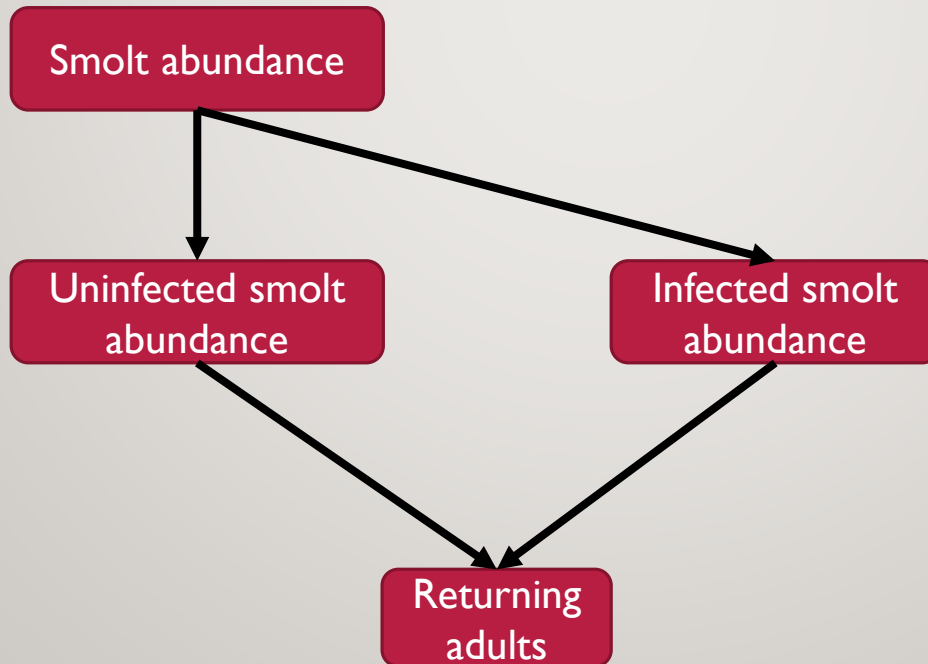
How do we achieve it?

Returning
adults

INTEGRATED APPROACH



INTEGRATED APPROACH



INTEGRATED APPROACH

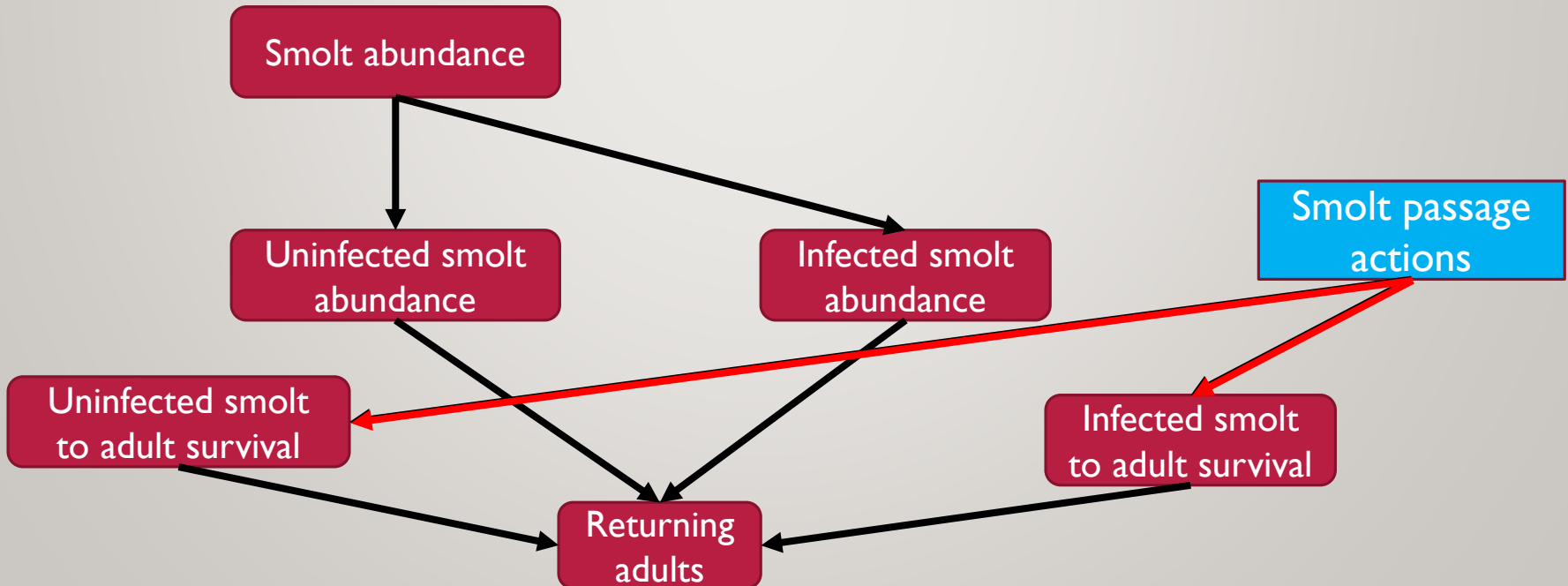
What is the best way to pass uninfected and infected juvenile?

Laboratory stress studies

Field evaluations

But... need a reliable supply of infected fish

>>> Infection experiments <<<



INTEGRATED APPROACH

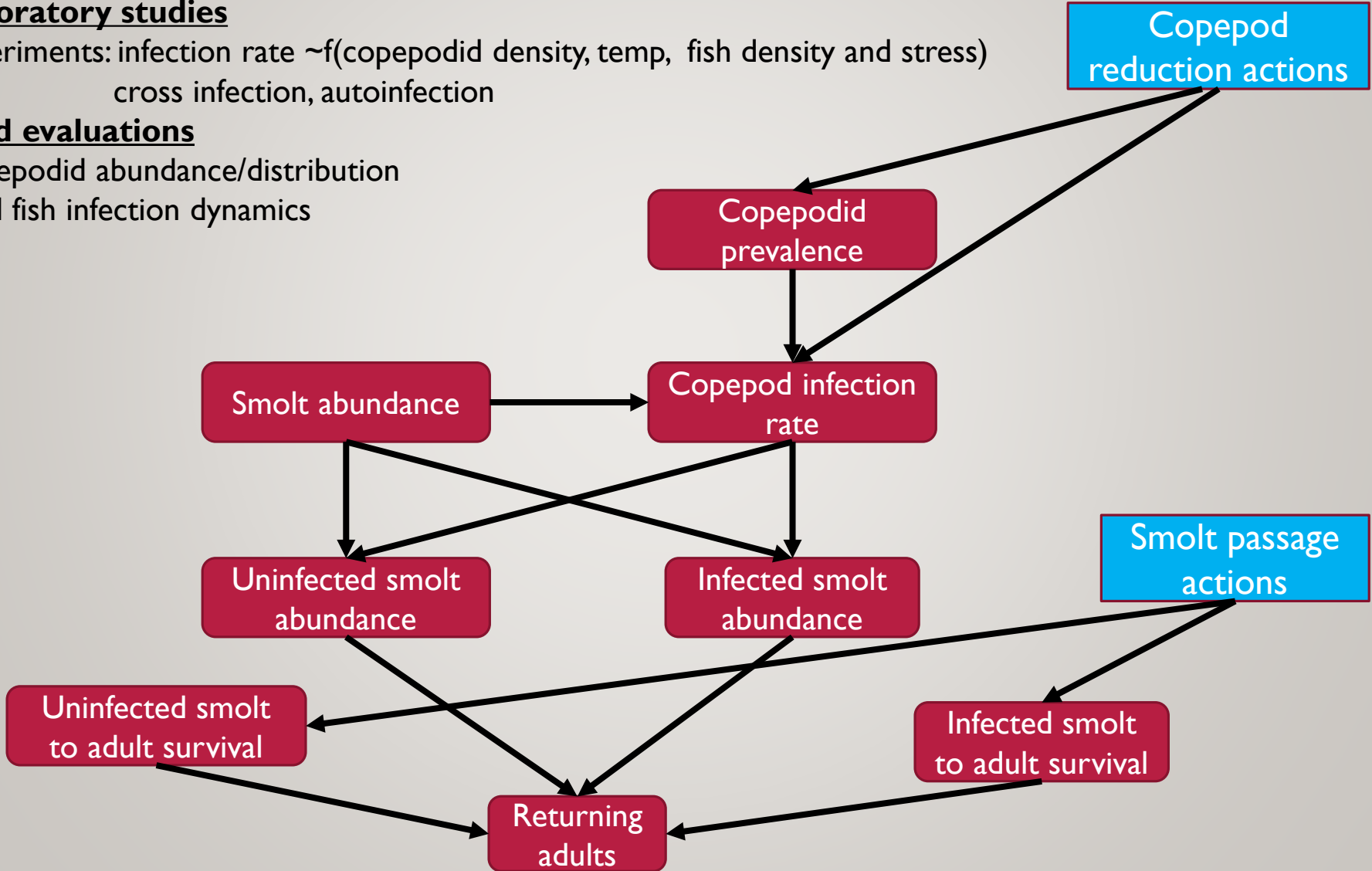
Why are juvenile Chinook so heavily infected?

Laboratory studies

Experiments: infection rate $\sim f(\text{copepodid density, temp, fish density and stress})$
cross infection, autoinfection

Field evaluations

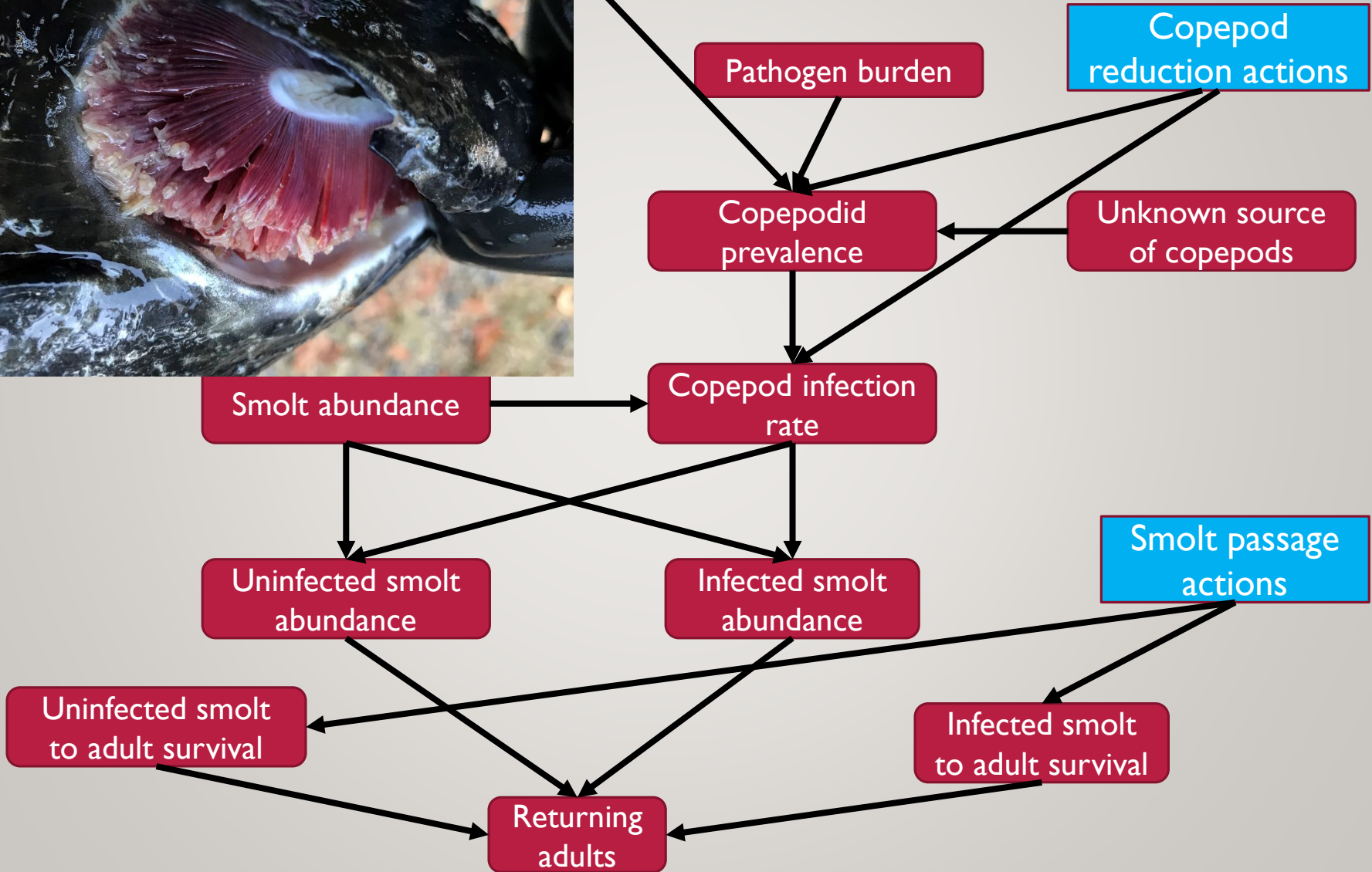
Copepodid abundance/distribution
Wild fish infection dynamics



INTEGRATED APPROACH



Plants? Infection?



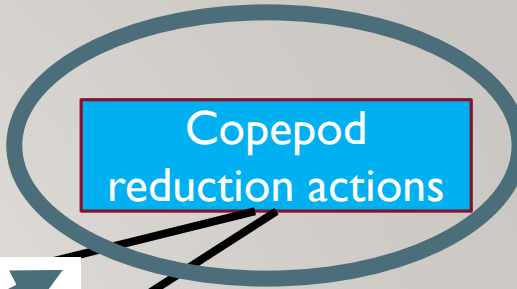
INTEGRATED APPROACH

Thermal exposure

No. outplants

Prespawn survival

Pathogen burden



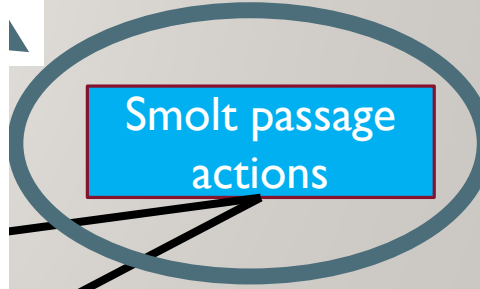
Copepod reduction actions

Source of copepods

Alternative management actions

Egg-to-smolt survival

Quantifiable fundamental objective



Smolt passage actions

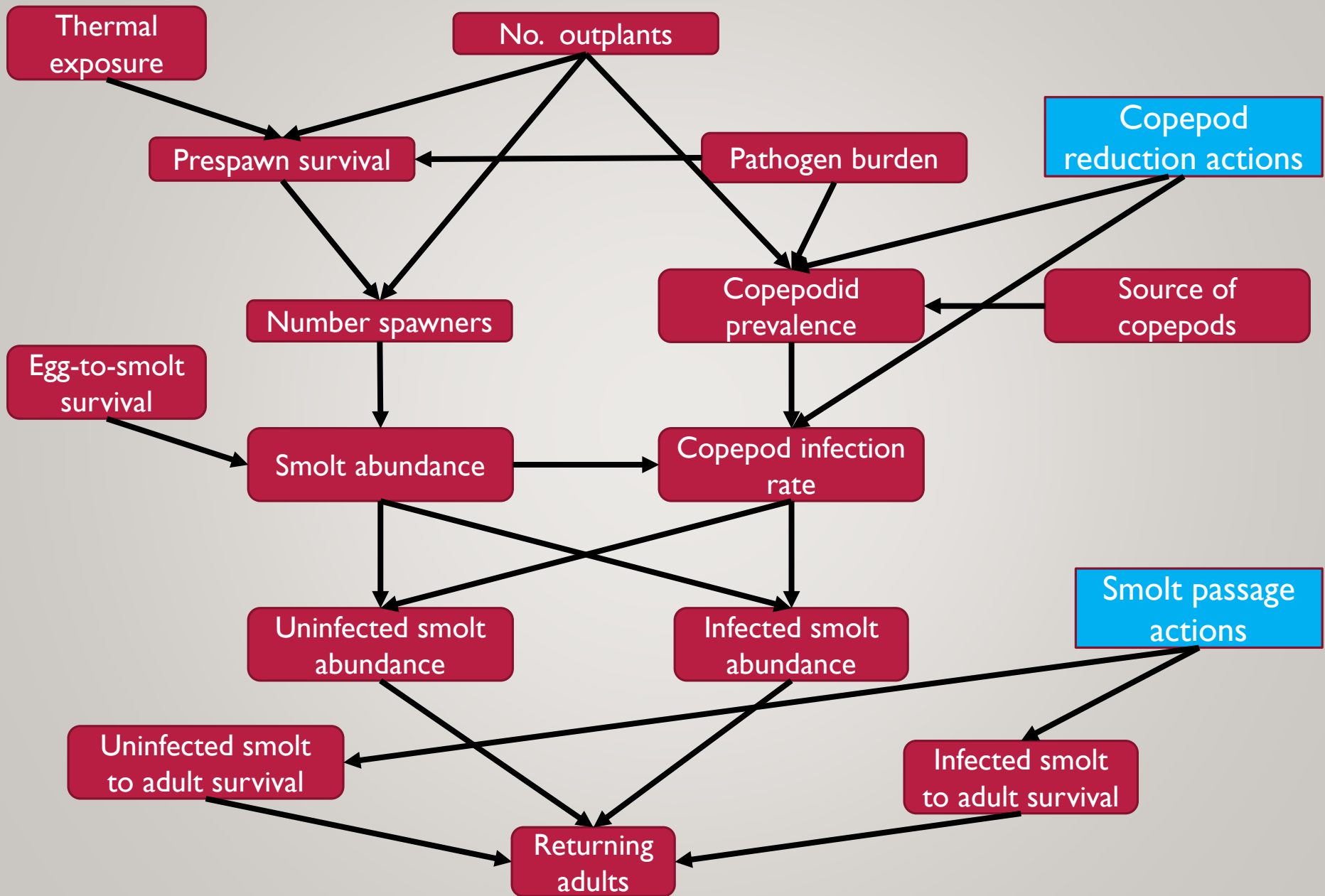
Uninfected to adult survival

Infected smolt to adult survival



Returning adults

BETA DECISION MODEL



PILOT INFECTION EXPERIMENTS

Two tank sizes- small (2' dia), large (3' dia) x 2 replicates

Copepodid density ~150-300 l

Water temperature- 12-13°C

Surrogate Chinook stocked at 1.6g/L (240 total fish)

20 infection events 16-Nov to 6 Dec

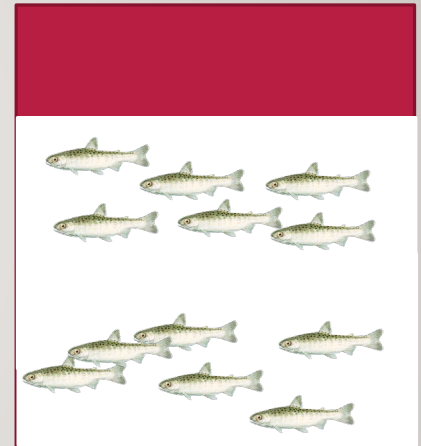
Infection event:

Lowered tank water

Introduced copepodids

Let stand 1 hr

Raised water level



ONGOING INFECTION EXPERIMENTS

Working hypotheses

1. Infection rate increases with increased stress
2. Infection rate and intensity increases with copepodid density
3. Infection rate increases with increased water temperature
4. Very high copepod infection intensities largely due to autoinfection
5. Very high copepod infection prevalence largely due to cross-infection

ONGOING INFECTION EXPERIMENTS

Treatments

Tank size (stress)- small (2' dia), large (3' dia)

Copepodid density- low (35-75 l), high (150-300 l)

Water temperature- cold (12-13°C), warm (15-16°C)

Fully factorial design $3^2 = 8$ trmts, 2 replicate tanks

Statistical power: main effects > 95%, interactions > 75%

Same infection protocol

Surrogate Chinook stocked at 1.6g/L (equal density)

Last infection dose Feb 4, ends week of March 24

INFECTION EXPERIMENTS

Cross infection

Large tanks- 3' dia

Cold water temperature- 12-13°C,

10 infected and 10 uninfected (ad clipped) surrogate fish

Hold fish minimum 6 weeks (complete copepod life cycle)

6 replicate tanks, 3 treatment x 3 control

Statistical precision: infection rate within 3% true value with
95% confidence

Initiated Feb 14, ends week of April 1

(Preliminary results)

ONGOING STRESS EXPERIMENTS

Replicate level of stress as trap and haul

Large tank 3' dia, 20 fish per tank

Cold water temperature 12-13°C

20 fish per tank

3 replicates control (uninfected) and infected ea

Initial experimental stressor- low level stressor

Crowd fish into center of tank

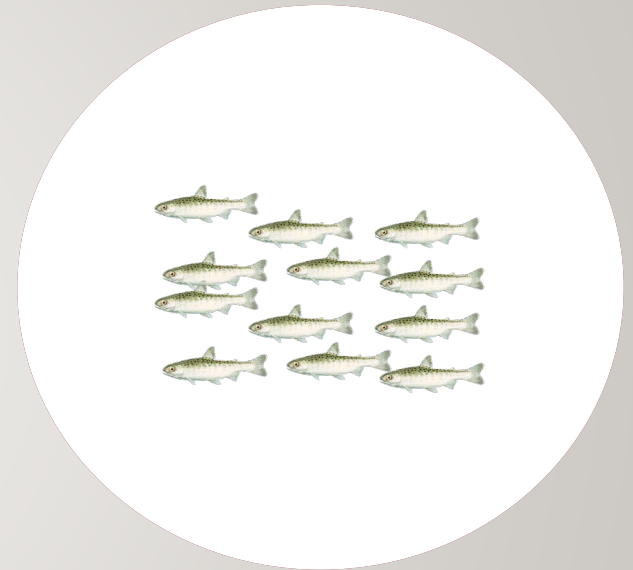
Hold 3 hr

Release fish from crowding

Sample cortisol at 1 hr, 6 hr, 24h, 2 weeks

Additional stresses as needed

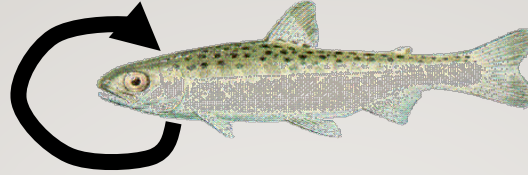
First test in about 2 weeks



UPCOMING EXPERIMENTS

Autoinfection

End of March



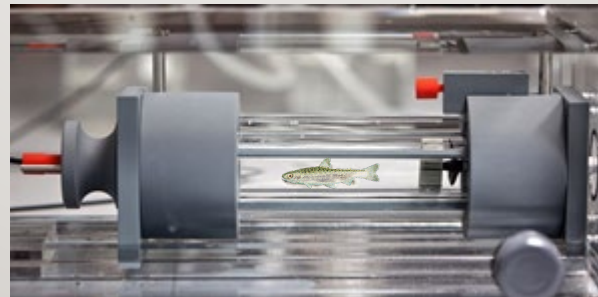
Saltwater challenge

End of March

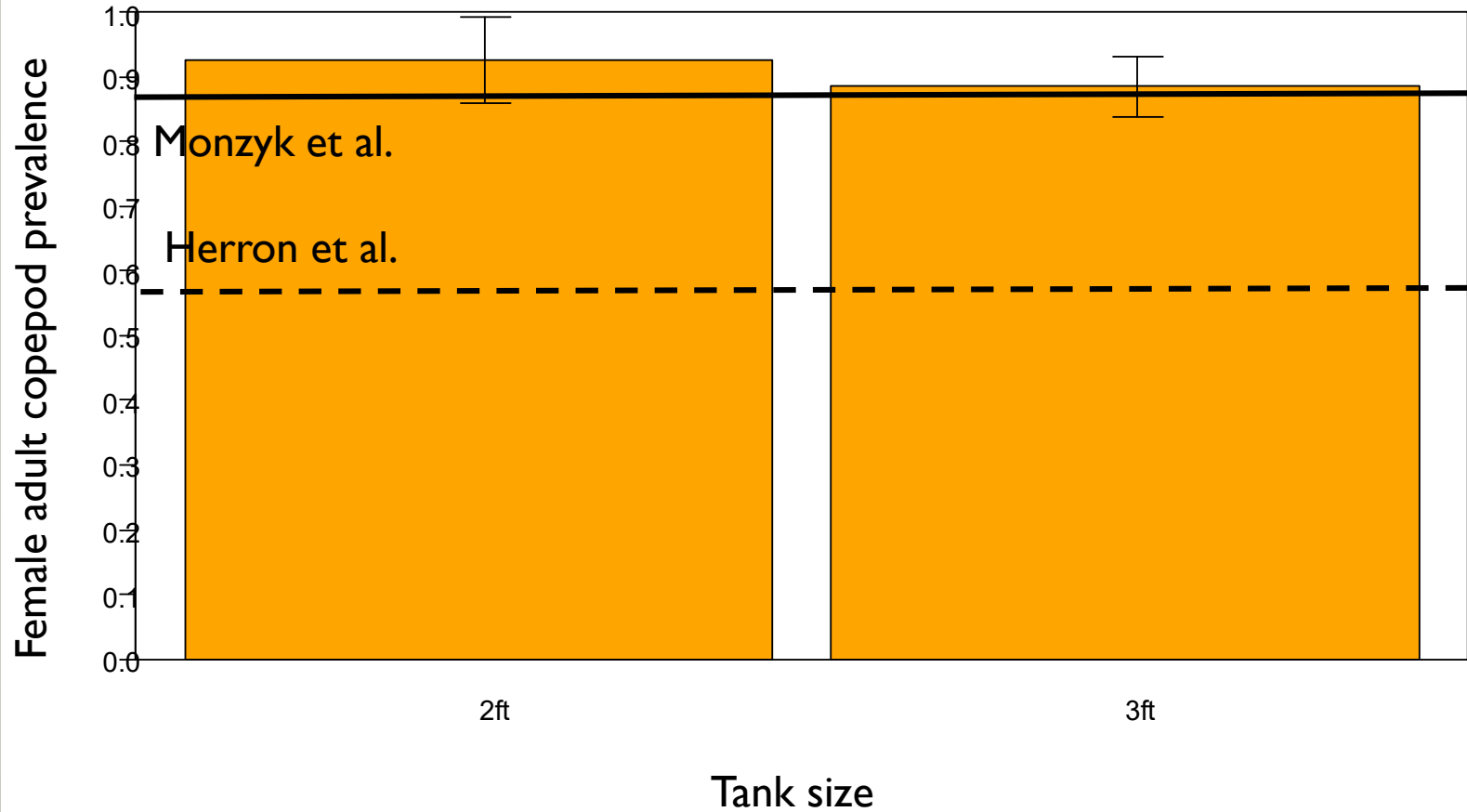


Swimming endurance

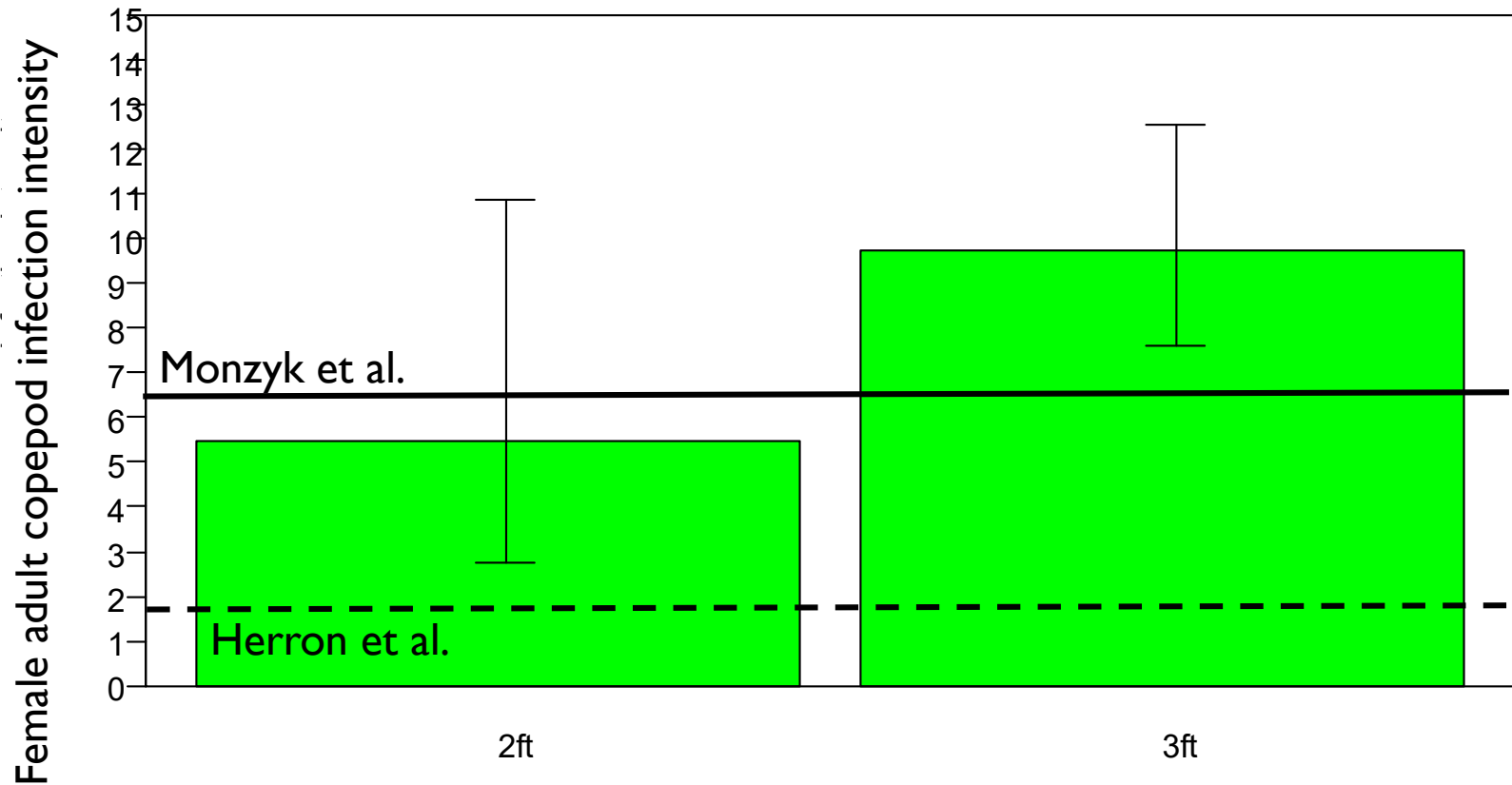
April-May



PRELIMINARY RESULTS



PRELIMINARY RESULTS



PRELIMINARY RESULTS

11 mortalities during pilot infection study

Mortalities had 2.87 times more copepods than fish that lived

73% attached to the gills or inside the operculum

Observed presence of pre-adult stages and significant gill damage

Observed evidence of cross or autoinfection in cross infection trials

Observed 2 mortalities transferring infected fish to tanks for stress evaluations, 0 for control fish

Infecting 200 rainbow trout consistent copepod source

CONCLUSIONS

We CAN infect juvenile Chinook salmon

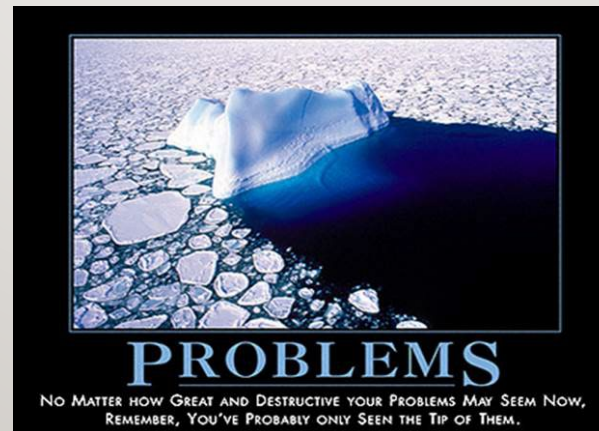
Prevalence and intensity equal to wild

Cross infection successful, large scale evaluations

Preliminary results ongoing studies April

Initiate structured decision making process

Infection intensity of adult female copepods are a poor indicator of the damage



ACKNOWLEDGEMENTS

Funding: USACE

ODFW

USACE

Oregon State University

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